

Received: September 12, 2023 Accepted: November 9, 2023 Published: April 24, 2024

Phosphorus-based Carrying Capacity of Aquaculture Sites in Lake Victoria, Kenya

©Sellu Mawundu^{1,4} ©Adiang' Evans^{1,2}, & ©Riziki Walumona Jacques^{1,3}

¹Department of Fisheries and Aquatic Science, University of Eldoret, P.O Box 1125-30100, Eldoret, Kenya

²Department of Agriculture and Environmental Studies, Ramogi Institute of Advanced Technology, Kisumu, Kenya

³Unité d'Enseignement et de Recherche en Hydrobiologie Appliquée (UERHA), Département de Biologie-Chimie, Institut Supérieur Pédagogique de Bukavu (ISP), Bukavu, D. R. Congo.

⁴Department of Aquaculture and Fisheries Management, Njala University PMB, Freetown, Sierra Leone

Abstract

Cage fish aquaculture is increasing rapidly in the African Great Lakes and reservoirs. In Lake Victoria, there is increasing number of cage fish farmers in the shallow bays of the lake. However, despite the proliferation of cages in the lake, there has been little regulation of cage aquaculture of the Nile tilapia (Oreochromis niloticus) in Lake Victoria leading to socio-economic conflicts and water quality concerns. In this study, we determined total phosphorous (TP) levels and derived water balance parameters, which were then used to compute TP assimilation capacity and fish production potentials for five sites within Kadimu Bay, one of the bays with active cage fish farming in Winam Gulf of Lake Victoria, Kenya. A mass-balance model was used to estimate the TP and fish carrying capacities of the fish cage sites in the bay. The model application is described in details in order to allow for replication in other African lakes and reservoirs, and in order to provide a framework for managing cage aquaculture and water quality. For all the sites, the TP assimilation capacity was exceeded by the TP released by the fish cages. Additionally, the maximum estimated fish production capacities were much less than the current fish production levels for all the sites. It is recommended that policies governing aquaculture production in the lake be reviewed based on these scientific findings and that the mass-balance model has potential applications for managing cage aquaculture in African lakes and reservoirs for sustainable fish production and water quality control.

Keywords: Mass-balance model, eutrophication, sustainability, Orechromis nilotic

Correspondence: nyamalahai38@gmail.com

Copyright © 2024 Sellu et al.. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY).

Competing interests: The authors have declared that no competing interests exist.

Introduction

Population growth and increase in per capita fish consumption demands water resources to be more efficient in terms of fish production on a global scale (Halwarth et al., 2007). The rising food demand is putting more pressure on wild fisheries resources (Worm et al., 2009) and cage fish farming is one of the alternatives increasingly being used to enhance fish production particularly in the tropics (FAO, 2010). Cage fish farming uses ecosystem services for the breakdown of organic matter, recycling of nutrients and supply of oxygen (Beveridge, 2004). However, a certain level of fish biomass may exceed the system capacity to function normally leading to a breakdown in ecosystem function through negative feedbacks such as eutrophication (Beveridge, 2004; Pillay, 2008; David et al., 2015), a process through which excess nutrients load especially phosphates and nitrates leads to increased algal production causing changes in ecosystem function and structure (Volleinweider, 1998). Cage aquaculture often results into water quality changes pollution through unconsumed, undigested and metabolic wastes from fish cages. Consequently, appropriate cage densities and fish production levels are required in order to provide optimum production without jeopardizing water quality and ecosystem function. The concept of ecological carrying capacity (Dillon and Rigler, 1975; Ross et al., 2013; David et al., 2015) that sets maximum limit for aquaculture production is a potential policy management tool sustainable cage aquaculture and

resilience of fresh water ecosystems to perturbations.

Aquaculture is increasing rapidly in African inland aquatic ecosystems (Njiru et al., 2004; Rothuis, 2014). In Lake Victoria for example, it is estimated that there are in excess of 4,000 cages on the Kenyan side of the lake (Njiru et al., 2018; Orina et al., 2018). Despite the popularity of the system, there have not been scientific studies to estimate the production carrying capacity of the lake as well as the ecological carrying capacity of cage sites for optimum aguaculture production. Such production should not jeopardize the ecological functions of the lake and should allow for sustainability and resilience of the ecosystem to perturbations. Sustainable cage culture production should focus on estimating the quantity of production that can be sustained by the environment without dramatic change in ecological processes, ecosystem services, species population and community structure (Beveridge, 2004; David et al., 2015). Massbalanced models are commonly being used to determine the optimum feeding regimes in cage aquaculture (Dillon and Rigler, 1975; Beveridge, 2004) and hence the carrying capacity of water bodies (Pulatsu, 2003; Ross et al., 2013; David et al., 2015). Phosphorus is a major limiting factor that phytoplankton productivity control especially in fresh water ecosystems (Vollenweider, 1998; Pillay, 2008) and therefore limits food chain productivity and often leads to eutrophication of aquatic systems when in excess. In this study, we used a mass-balanced model to

estimate the total phosphorous and fish production carrying capacity of five cage sites for Nile tilapia (*Oreochromis niloticus*) production in Kadimu Bay, one of the five active aquaculture sites in Lake Victoria, Kenya. The applied methodology and results should provide a pioneering benchmark for managing aquaculture production in African lakes and reservoirs.

Materials and Methods

Study area

The study was conducted within Kadimu Bay (Figure 1), one of the five main bays of Lake Victoria, Kenya. The bay is situated between latitude 0° 5′ 10″ S and longitude 34° 6′4″E and lies within the tropical monsoon climate (Kottek, 2006). The depth range of the bay is between 3 to 16 m, about 947 km² in area and spans a

distance of about 4.3 km (Calamari et al., 1995). Its shallow and sheltered nature makes it suitable for cage fish farming. The larger Winam Gulf that forms the Kenyan side of Lake Victoria lies on latitude 0° 14'24"S and longitude 34° 34' 48"E (Geheb, 1977). The annual average precipitation over the lake is about 1 300 mm with average annual temperature of about 22.9°C (Masongo et al., 2005). Most of the sheltered bays in Lake Victoria have cage fish farming as a recent introduction (Aura et al., 2018). The cages are medium sized floating types with typical dimensions of 7 x 8 x 9 m. The main cultured species in the lake is the Nile tilapia (Oreochromis *niloticus*) and the culture practice is mainly semi-intensive (Njiru et al., 2018). The fish are fed with commercial pellets supplemented with farmer-formulated feeds comprising of the fresh water shrimp (Caridina spp.).

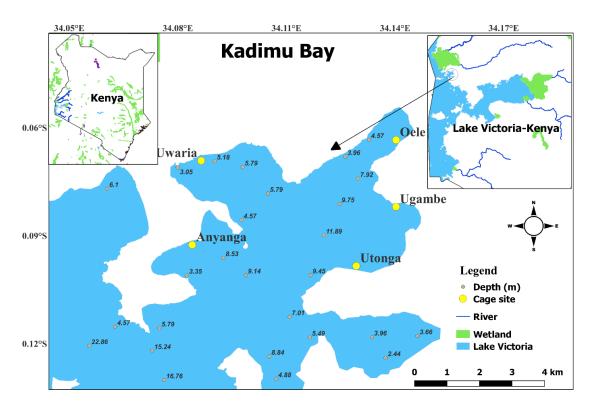


Figure 1: Map of Lake Victoria showing sampling sites: Anyanga, Uwaria, Oele, ugambe and, Utonga in the Kadimu Bay of Winam Gulf, Lake Victoria, Kenya. (Modified from KMFRI).

Sampling

Sampling for physico-chemical parameters and nutrient load conducted at five fish cage sites (Anyanga, Uwaria, Oele, Ugambe and Utonga) within the Kadimu Bay (Figure. 1). The five cage sites have an average depth of 7.098 m and are separated by an average distance of 0.5 Km. Each cage site is managed under a different beach management group. The sites were selected because they had ongoing cage fish farming activities, were easily accessible and met most of the site selection criteria (Pillay, 2005). Sampling for water quality variables was conducted at a frequency of three-times in a month for a period of ten months extending from January to October, 2021. On each sampling occasion, three replicates of water samples were collected randomly at the sites using labeled water bottles. The samples were placed in a cooler box at a temperature of approximately 4°C and transported to the Kenya Marine and Fisheries Research Institute (KMFRI) laboratory for analysis of total phosphorus concentrations. Water transparency was measured in situ with a Secchi disk of about 20 cm diameter (Bartram and Balance, 1996).

Analytical procedures

The UV Spectrophotometer (Genesys 10S Vis SN- 2F1N308001), was used for analysis of chlorophyll-a, total phosphorus (TP), and total nitrogen (TN) following the methods in APHA (2005). The partitioning of chlorophyll-a was done by the sonication technique (APHA, 2005) and the concentration determined by the Lorenzen equation (APHA, 2005) through the application of absorbance readings from the UV Spectrophotometer. ascorbic reduction technique and the diazotization procedure were used to determine TP and TN concentrations using unfiltered water samples (APHA, 2005). A total of eighteen water samples were analyzed per site.

Calculation of Phosphorus-based carrying capacity

Phosphorus is a critical element needed by fish for optimum growth and development (Pillay, 2005). It is often a limiting nutrient in freshwater bodies that controls phytoplankton production and together with light, optimizes productivity of aquatic ecosystems (Beveridge, 2004). Over supply phosphorus leads to non-linear feedbacks those results into negative responses like eutrophication and water quality deterioration (Volleinweider, Beveridge, 2004; Pillay, 2005). Phosphorusbased carrying capacity (mg m³) is the TP assimilation potential of an area that will not disrupt the ecological functioning of the area (Beveridge, 2004). Total phosphorus load of a site is simply quantified by multiplying the site area by unit concentration of TP in the water. The TP assimilation load of the five cage sites within the study area were estimated following the mass-balance equation modified from Dillon and Rigler (1975) and as applied by Beveridge (2004) and Pulatsu (2003) as:

Lfish =
$$\frac{\Delta[P]Z\emptyset}{1-Rfish}$$
(1)

Where:

Lfish = Cage site carrying capacity (maximum assimilation load) of total phosphorus (mg m³ year-¹).

 Δ [P] (mg m⁻³) = Total phosphorus allocation load, obtained from the difference between measured phosphorus [P]_c and the maximum allowable phosphorus load [P]_i for fish culture as:

$$\Delta[P] = [P]i - [P]c \dots (2)$$

Z (m) = Average depth of a cage site

 \emptyset (year⁻¹) =Flushing coefficient, a measure of the water replacement rate at a site.

Estimated from average water debit (Q, m³ year⁻¹) and water volume at sites (V, m³) as:

$$\emptyset = Q/V$$
(3)

Where.

R_{fish} = Proportion of total phosphorus left in sediments as a result of feeding fish in the cages and from fish waste matter estimated according to Beveridge (2004) and Pulatsu (2003) as:

$$Rfish = X + [(1 - X)]R$$
(4)

Where,

X: the net proportion of total phosphorus lost permanently to the sediments as a result of solid deposition and is estimated to be ranging between 0.4-0.5 (Pulatsu, 2003).

R: Phosphorus retention coefficient given by:

$$R = \frac{1}{1 + 0.747 \text{ } \% \text{ } ^{\circ} 0.507 \text{ } \dots (5)}$$

The effective quantity of total phosphorus discharged in the environment from fish aquaculture waste (Pe) is directly proportional to fish production and is given by the equation (Beveridge, 2004):

$$Pe = (Pfeed \ x \ FCR) - Pfish.....(6)$$

Where:

Pe = TP released into the environment from a ton of fish produced (kg of TP per ton of fish production).

FCR = Feed conversion ratio generated from the ratio of feed given and the amount of fish produced.

 $P_{feed} = TP \text{ in fish feeds (kg ton}^{-1})$

P_{fish} = TP incorporated into the body of whole fish (kg ton⁻¹)

Data sources for calculation of Phosphorus-based carrying capacity

The variables for the estimation of phosphorus-based carrying capacity per site using the mass-balanced model (equation 1) were derived through fieldwork activities and from the published and grey literature. The surface area and water volume at each cage site were estimated by KMFRI using the planimetry method. The average depth (Z, m) of cage sites were estimated using a depth finder. The average water debit (Q, m³ year⁻¹) for the cage sites used to derived the flushing rate (\emptyset) in equation 3 was determined from the average of published data on the lake's water balance parameters for comparable sites and included; long-term average inflow from catchment surface runoff (Ad..r), cage site surface area (A), Precipitation into the lake (Pr), and Evaporation from the lake (Ev) (Table 1). The variables were incorporated in the equation below as (Dillon and Rigler, 1975; Shoji, 2009);

$$Q = Ad.r + A (Pr - Ev) \dots (7)$$

Analysis of TP in feed (Pfeed) and TP in the body of whole fish (Pfish) (equation 6) was done at the KMFRI laboratory. Consequently, the TP in feed used by cage fish farmers was estimated to be about 21.75 kg ton⁻¹, while TP in the body of whole tilapia was estimated at about 3.5 kg ton⁻¹.

Table 1: Published estimates of the mean annual water balance parameters of Lake Victoria, and their reference sources. Adopted from Xungang and Nicholson (1998)

		Evaporatio	Tributary	
	Rainfall Over lake	n	inflow	
Period	(mm/year)	(mm/year)	(mm/year)	Reference source
	1420	1350	230	Hurst (1952)
	1145	1130	237.5	
4025 4050	4.620	4522	260	1.0.1(1070)
1925-1959	1630	1523	260	de Baulny and Baker (1970)
				Hastenrath and
	1650	1500	250	Kutzbach(1983)
	1636	1459.5	238	WMO (1974, 1981)
	1450	1370	260	Spigel and Coulter (1996)
1956-1978	1810	1593	343	Howell et al (1988)
1945-1984	1645	1470	0	Flohn and Burkhardt (1985)
1950-1979	1660	1590	420	Kite (1982)
1970-1974	1850	1595	343	Piper et al (1986)
1956-1978	1476	1401	241	Balek (1977)
Mean	1579.27	1452.86	256.59	

Results

Phosphorus-based carrying capacity

The mass-balanced model in equation (1) was applied to derive the total phosphorus (TP) assimilation capacity for the five sites. An example of how TP assimilation capacity was derived for each of the five cage sites

in Kadimu Bay is shown for Anyanga site (Figure. 1) as follows:

For Anyanga site, the TP assimilation carrying capacity (amount of TP the area can hold as a result of cage fish farming), L fish, in equation (1) was derived from the following steps:

Table 2: Datasets for calculation of total phosphorus assimilation carrying capacity for cage sites in the Kadimu Bay, Lake Victoria, Kenya

	Surface area	Dept	Volume (V)	Water Debit (Q)	Flushing rate
Site	(m²)	h (m)	(m³)	(m³/year)	(Ø=Q/V) (year ⁻¹)
Anyanga	617509	7.01	4328738.09	78059569.28	18.03
Oele	22458	4.27	95895.66	2839172.37	29.61
Ugambe	9250	8.84	81770	1169549.09	14.30
		10.6			
Utonga	29122	7	310731.74	3681568.61	11.85
Uwaria	137022	4.7	644003.4	173321207.61	26.89

The TP allocation load Δ [P] was derived as 13.44 mg m⁻³ from the difference between maximum allowable TP allocation load of 100 mg m⁻³

recommended for Lake Victoria (Kenya) cage aquaculture by the Kenyan Marine and Fisheries Research Institute (Aura, 2020), and the highest measured field value of 86.56 mg m⁻³ at the site. The highest field values of TP at sites were used

in order to provide precautionary estimates of TP assimilation carrying capacities.

Given the average depth (Z) of 7.01 m for Anyanga site (Table 2) and flushing rate (\emptyset , Equation 3) of 18.03 year⁻¹ derived from water debit (Q) of 78 059 569.28 m³ year⁻¹ from equation (3) and water volume (V) of 4 328 738.09 m³ (Tables 1 and 2), the phosphorus retention coefficient (R) was derived as 0.24 from equation (5).

Consequently, R fish (equation 4), the proportion of TP left in sediment was derived as 0.62 taking X as 0.5. Therefore, the amount of TP that can be carried by the lake at Anyanga site (Lfish) was estimated by substituting the above variables into equation (1) as 4.47 g m⁻² year⁻¹ (Table 3). This was multiplied by the cage site surface area of 617 509 m² to derived the site overall TP assimilation carrying capacity as 2 760 265.23 g year⁻¹ or equivalent to 2 760.27 kg year⁻¹ (Table 4).

Table 3: Total Phosphorus (TP) assimilation carrying capacity, fish production carrying capacity and hydrological parameters of cage sites within Kadimu Bay, Lake Victoria, Kenya

Site	TPmax (mg/m³)	ΔP(mg m-³)	Z(m)	Ø(year ¹)	R	R_{Fish}	Lfish (g/m²/year)
Anyanga	86.56	13.44	7.01	18.03	0.24	0.62	4.47
						0.59	
Oele	96.61	3.39	4.27	29.61	0.19	5	1.058
Ugambe	97.38	2.62	8.84	14.3	0.26	0.63	0.895
			10.6				
Utonga	82.93	17.07	7	11.85	0.28	0.64	5.995
Uwaria	55.16	44.84	4.7	26.87	0.2	0.6	14.167
Kadimu			7.09		0.23	0.61	
Bay	83.73	16.27	8	20.132	4	7	6.07

All notations are as explained in the text.

Following equation (6) for the amount of TP generated per ton of fish produced, and given the TP in feed (P_{feed}) as 21.75 kg ton⁻¹, FCR value of 1.4 and TP in the body of whole fish (P_{fish}) as 3.5 kg ton⁻¹ the TP released in the environment per ton of fish produced (Pe) at the site was estimated at 26.95 kg ton⁻¹. Consequently, the total carrying capacity for Anyanga site based on fish production is derived as 102 421.72 kg year⁻¹ (102.42 MT year⁻¹ Table 4) from the ratio of TP assimilation capacity for the site (2 760.27 kg year⁻¹) and the quantity of phosphorus released per ton of fish produced (26.95 kg).

From consultations and the literature (Orina et al., 2018), the annual

cage fish production at Anyanga site averages 6 320 MT year⁻¹ (Table 4). This production therefore releases 170 324 kg year⁻¹ (Pe x fish production) of TP to the environment at the site. This value is several magnitudes higher than the estimated TP carrying capacity at Anyanga site of 2 760.27 kg year⁻¹

The TP assimilation capacity for each of the other four sites, the TP released by current fish production at sites, the current fish production at sites and the maximum potential fish production at sites were calculated in the same computational procedure and are shown in Table 4.

Table 4: TP carrying capacity, TP release by cu	rrent fish production, Maximum potential fish
production and current fish production by cage	e site in the Kadimu Bay, Lake Victoria, Kenya.

Site	TP carrying capacity (kg/year)	TP released by current fish production (kg/year)	Maximum potential fish production (ton/year)	Current fish production (ton/year)
Anyanga	2,760.270	170,324	102.420	6,320
Oele	23.761	113,190	0.882	4,200
Ugambe	8.279	32,340	0.307	1,200
Utonga	174.586	6,468	6.478	240
Uwaria	1,941.191	21,560	72.029	800
Kadimu Bay	4,949.241	343,882	183.645	12,760

The estimated cage site fish production potential within the studied bay (Table 4) ranged from a minimum of 0.307 MT year⁻¹ at the Ugambe site to a maximum of 102.42 MT year-1 at the Anyanga site showing variations in production potential at small spatialscales. The other sites also had variable fish production potential in the bay that varied from a relatively high yield at Uwaria (72.03 MT year⁻¹), low values for Utonga (6.48 MT year⁻¹), Ugambe (0.31 MT year⁻¹), and Oele (0.88 MT year⁻¹). The estimated potential maximum fish yield for all the five cage sites were orders of magnitude lower than the current harvests by the farmers (Table 4).

The estimated TP released to the environment by current fish production (Pe x current fish production) is greater than the TP that can be accommodated by sites (Lfish) for all the sites, indicating that the TP assimilation capacity at the cage sites has been exceeded (Table 4). Hence, the potential for more fish production at the cage sites in the Kadimu Bay, including the TP that can be accommodated by these cages sites (Lfish), indicates that both the maximum potential fish production and TP carrying capacity for the bay have been exceeded (Table 4). The TP released by current fish production in the Kadimu Bay

(Table 4) is a lot more than the TP that can be accommodated by the bay. The current cage fish production in the Kadimu Bay (Table 4) is above the estimated maximum potential cage fish production of the bay following the mass-balanced model used in this paper.

Discussions

This paper pioneers the use of nutrient analysis, especially TP, in setting the limit of aguaculture production in African lakes and reservoirs. The mass-balance method can provide a useful tool for managing cage aguaculture in lakes and reservoirs (David et al., 2015; Pulatsu, 2003). However, the accuracy of the TP based mass-balanced model applied in this study, will depend on the availability of accurate and recent data on the water balance parameters of aquatic systems, something which is lacking for most African lakes and reservoirs (Roberts and Zohary, 2018; Plisnier et al., 2022). In this study, we relied on historical water balance data for the Winam Gulf of Lake Victoria due to the unavailability of recent datasets. The estimates provided in this paper will therefore require validation as more physical and limnological data become available on Lake Victoria. Nonetheless, the

paper has delved into methodological and computational details to allow replication of the method as a policy tool for managing aquaculture in the African Great lakes and reservoirs.

The estimated allowable TP level for the studied bay is exceeded by the current TP released into the environment as a result of aquaculture activities. Excess TP can lead into increased algal biomass production through eutrophication (Pillay, 2008) and water quality deterioration. Some studies on the Winam Gulf have found TP levels not to have reached the eutrophication thresholds (Gikumu-Njuru et al., 2021) save for occasional seasonal increases in TP loading caused by agricultural run-off leading to localized eutrophication (Ledang et al., 2020). TP levels in this study ranged between 85.11 and 140.02 mg m⁻³ and are comparable to those of other studies in Lake Victoria (Gikuma-Njuru et al., 2021) and provided a moderate Eutrophic state (Sellu unpublished data). However, human activities including deforestation, urbanization and agricultural production have the potential to increase nutrient load into Lake Victoria (Ogutu-Ohwayo, 1990; Roberts and Zohary, 2018) and could exacerbate the enrichment caused by aquaculture in the lake.

The maximum TP assimilation load for the studied sites has a direct correlation with the depth, flushing rates and allowable TP as per the mass-balanced model (sensu Dillon and Rigler, 1975). Except for the depth variable, the other parameters will require more validation in future studies. For example, the accuracy of TP allocation load (Δ [P]) based on maximum TP concentration of 100 mg m⁻³ (Aura, 2020) will affect the maximum TP an area can hold from aquaculture and the potential fish production based on the model. The maximum allowable TP value in lakes varies between jurisdictions with a

recorded maximum of 300 mg m⁻³ in some waters (Pulatsu, 2003) and low values of up to 30 mg m⁻³ in reservoirs (David et al., 2015). There is no clear policy on the maximum allowable TP values in Lake Victoria and hence requiring more validation of the proposals by KMFRI (Aura, 2020) in future studies.

The TP based mass-balanced model used in this study suggests the TP and Nile tilapia carrying capacity for the sites in the bay have been surpassed by the current fish production by farmers in the bay. These estimates are based on a precautionary approach but are useful starting guidelines for the management of Lake Vitoria cage aquaculture. There has not been a clear policy guideline on the number of cages allowed in Lake Victoria, leading to conflict between the farmers and wild stock fisheries (Njiru et al., 2018). Determination of carrying capacities for fish production and TP loads in the lake's shallow bays is important for a planned cage aquaculture enterprise including the setting up of sites for "aquaculture parks" (sensu David et al., 2015). TP addition by the cage aquaculture in the lake is likely to synergize with run-off inputs from agricultural activities around the lake hence the novelty of precautionary approach used to estimate TP allocation levels in this study and others (David et al., 2015; Mahamudi, 2019).

Conclusions and recommendations

This study provides a pioneering policy tool for managing aquaculture development in Lake Victoria and other African water bodies for sustainable livelihoods and ecosystem functioning. The mass-balanced model outputs used in this study will require re-parameterization as more data become available but are an important starting guideline for science-

based management of aquaculture in the lake. Scientific effort is therefore required in order to generate recent data on water balance parameters for the lake, in addition to other limnological variables necessary to fit the model.

Acknowledgements

The authors would like to sincerely thank the Beach Management Units (BMUs) at Kadimu Bay for granting access to the cage sites for data collection. Special thanks and appreciations to the technical team of the KMFRI, Kisumu, Kenya. The Carnegie Cooperation of New York (CCNY) for providing financial support through the Regional Universities Forum for Capacity Agriculture (RUFORUM) Graduate Teaching Assistant (GTA) program.

Funding

This project was funded by the Carnegie Cooperation of New York (CCNY) through Regional Universities Forum for Capacity Building in Agriculture (RUFORUM) Graduate Teaching Assistant (GTA) program.

References

- APHA, (2005). American Public Health Association Standard methods for the examination of water and waste water, 21st Ed. Washington, D.C., APHA- AWWA –WEF, ISBN 0875530478.
- Aura, C. M., (2020). A brief on fish cage farming in Lake Victoria, Kenya as guidance on decision making for policy direction, Kenya Marine and Fisheries Research Institute (KMFRI), 9 pp.
- Aura, C. M., Musa, S., Yongo, E., Okechi, J.K., Njiru, J.M, Ogari, Z., Wanyama, R., Charo-Karisa, H., Mbugua, H., Kidera, S., Ombwa,V., Oucho, J.A., (2018). Integration of mapping and socio-economic status of cage culture: Towards balancing Lake-use and culture Fisheries in Lake Victoria Kenya, journal of Aquaculture research 49(1), 532-545. DOI: 10 1111/are. 13484.

Balek, J., (1977). Hydrology and Water Resources of Tropical Africa. Elsevier, Amsterdam.

- Bartram, J., Balance, R., (1996). Water quality monitoring: A practical guide to the design and implementation of freshwater quality studies and monitoring programmes UNEP/UNESCO.
- Beveridge, M. C. M., (2004). Cage aquaculture, 3rd ed. Wiley-Blackwell, Oxford, UK, pp 368.
- Byron, C., Jason, L., Costa-Pierce, B., Bengton, D., (2011). Calculating ecological carrying capacity of Shellfish aquaculture using mass-balance modelling, Aquaculture 222, 1743-1755. DOI: 10.1016/j.ecolmodel.2011.03.010.
- Calamari, D., Akech, M.O., Ochumba, O., (1995).

 Pollution of Winam Gulf, Lake Victoria
 Kenya: A case study for preliminary risk
 assessment. Lake and Reservoir Research
 and management 1, 89-106.
- David, G.S., Carvalho, E.D., Lemos, D., Silveira, A.N., Dall'Aglio-Sobrinho, M., (2015). Ecological carrying capacity for intensive tilapia *Oreochromis niloticus* cage culture in large hydroelectrical reservoir in South-Eastern Brazil. Aquaculture Engineering 66, 30-40. DOI:org/10.1016/j.aquaeng.2015.02.003
- de Baulny, H.L., Baker, D., (1970). The water balance of Lake Victoria. Tech Note Water Development Department, Republic of Uganda.
- Dillon, P.J., Rigler, F.H., (1975). A simple method for predicting the carrying capacity of lake trophic status. Journal of fisheries research Board Canada 32, 1519-1531.
- FAO, (2010). The state of world fisheries and aquaculture, Rome, Italy. 354 pp
- Flohn, H., Burkbardt, T., (1985). Nile runoff at Aswan and Lake Victoria, a case of a discontinuous climate time series. Z Gletscherk Glazialgeol. 21, 125-130.
- Geheb, K., (1977). The regulation and the regulated fisheries management options and dynamics in Kenya's Lake Victoria, fisheries (PhD thesis), University of Sussex.
- Gikuma-Njuru, P., Rutagemwa, D.K., Mugidde, R., Hecky, R.E., Mwebaza-Ndawula, L., Mwirigi, P.M., Abuodha, J.O.Z., Waya, R.K., Matovu, A., Kinobe, J., (2021). Eutrophication of the Lake Victoria ecosystem. Lake Victoria environment management project (LVEMP) http://hdl.handle.net/1834/6902.
- Halwarth, M., Soto, D., Arthur, J.R., (2007). Cage Aquaculture-Regional review and Global Overview pp 1-16 FAO Fisheries Technical paper No. 498 Rome, FAO.

Hastenrath, S.L. Kutzbach, J. E., (1983). Paleoclimate estimates from water and energy budgets of East African Lakes. Quatern, Res. 19, 141-153.

- Hecky, R.E., Campbell, P., Hendzel, L.L., (1993). The stoichiometry of carbon, nitrogen and phosphorus in particulate matter of Lakes and Oceans. Limnol.Oceanogr. 38 (4), 709-724.
- Howell, P.P, Lock, M., Cobb, S., (1988). The jonglei canal: Impact and opportunity, Cambridge University press. Cambridge.
- Hurst, H.E., (1952). The Nile basin constable, London ISSN: 0262-6667 Hydrological Science Journal 61(9), 1557-1570.
- Joseph, S., (2009). Open sea cage culture: Carrying Capacity and stocking in the grow out system. National Training on cage culture of seabass held at CMFRI, Kochi from 14-23 December, 2009.
- Kite, G.W., (1982). Analysis of Lake Victoria levels. Hydrol. Sci. J. 27(2), 99-110.
- Kottek, M., Grieser, J., Beck, C., Rudolf, B., Rubel, F., (2006). World map of the Koppen-Geiger climate classification updated. Meteorology 15, 259-263. DOI: 10.1127/0941-2948/2006/0130.
- Ledang, A.B., Odermatt, N.D., Brockmann, O., (2020).

 Traces of Eutrophication in Winam Gulf
 Lake Victoria. Inland waters Diversity
 report.
- Mahmudi, M., Lusiana, E.D., Arsad, S., Buwono, N.R., Darmawan, A., Nisya, T.W., Gurinda, G.A., (2019). A study on phosphorus based carrying capacity and trophic status index of floating cages area in Rana Grati. AACL Bioflux. 12: No 5.
- Masongo. O., Mwirigi, P., Okungu J., Osio, J., Abuodha, J.O.Z., Hecky, R.E., (2005). Sedimentation in Lake Victoria and the Winam Gulf Book Chapter 9, 211-265. Url http://hdlhandle.net/1834/7141.
- Njiru, J.M., Aura, C.M., Okechi, J.K., (2018). Cage fish culture in Lake Victoria: A boon or a disaster in waiting? Fisheries Management and Ecology 26(5), 426-434. DOI: 10.1111/fme.12283.
- Njiru, M., Okeyo-Owour, J.B., Muchiri, M., Cowx, I.G., (2004). Shift in the food of Nile tilapia Oreochromis niloticus. African Journal of Ecology 42, 163-170.
- Oguto-Ohwayo., (1990). The diversity and abundance of zooplankton in Lake Victoria and Kyoga basins and their relationship to fish production. National Fisheries Resources Research Institute (NaFIRRI). http://hdl.handle.net/1834/35207.

Orina, P.S., Ogello, E., Kembenya, E., Githukia, C., Musa, S., Ombwa, V., Mwainge, V.M., Abwao, J., Ondiba, R.N., Okechi, J. K., (2018). State of cage culture in Lake Victoria, Kenya. https://www.researchgate.net/publicatio n/333805185

- Pillay, T. V. R. Kutty, M. N., (2005). Cabdirect.org [Book] Aquaculture: Principles and Practices.
- Pillay, T.V.R., (2008). [Book] Aquaculture and the environment second edition.
- Piper, B.S., Plinston, D.T. Sutcliffe, J.V., (1986). The water balance of Lake Victoria Hydrol. Sci. J. 31(1), 25-37.
- Plisnier, P.D., Kayanda, R., MacIntyre, S., Obiero, K., Okello, W., Vodacek, A., Cocquyt, C., Abegaz, H., Achieng, A., Akonkwa, B., Albrecht, C., Balagizi, C., Barasa, J., Bashonga, R.A., Bishobibiri A.B., Bootsma, H., Borges, A.V., Chavula, G., Dadi, T., De Keyzer, E.L.R., Doran, P.J., Gabagambi, N., Gatare, R., Gemmell, A., Getahun, A., Haambiya, L.H., Higgins, S.N., Hyangya, B.L., Ir vine, K., Isumbisho, M., Jonasse, C., Katongo, C., Katsev, S., Keyombe, J., Kimirei, I., Kisekelwa, T., Kishe, M., Koding, S.O.A., Kolding, J., Kraemer, B.M., Limbu, P., Lomodei, E., Mahongo, S.B., Malala, J., Mbabazi, S., Masilya, P.M., McCandless, M., Medard, M., Ajode, Z.M., Mrosso, H.D., E.R., Mudakikwa, Mulimbwa, N.; Mushagalusa, D., Muvundja, F.A., Nankabirwa, A., Nahimana, D., Ngatunga, B.P., Ngochera, M., Nicholson, S., Nshombo, M., Ntakimazi, G., Nyamweya, C., Nyeko, J.I., Olago, D., Olbamo, T., O'Reilly, C.M., Pasche, N., Phiri, H., Raasakka, N., Salyani, A., Sibomana, C., Silsbe, G.M., Smith, S., Sterner, R.W., Thiery, W., Tuyisenge, J., Van der Knaap, M., Van Steenberge, M., van Zwieten, P.A.M., Verheyen, E., Wakjira, M., Walakira, J., Wembo, O.N., Lawrence, T., (2022). Need for harmonized long-term multi-lake monitoring of African Great Lakes, Journal of Great Lakes Research, https://DOI:.org/10.1016/j.jglr.2022.01.01
- Pulatsu, S., (2003). The application of phosphorus budget modle estimating the carrying capacity of Kesikkopru Dam Lake. Turkish Journal of Vetenary and Animal Science 27 (5), 1127-1130.
- Roberts, R.D., Zohary, T., (2018). Limnology and future of African Inland waters. Inland waters 8(4), 399-412.

- Ross, L.G., Telfer, T.C., Falconer, L., Soto, D., Aguilar-Manjarrez, J., Asmah, R., Bermudez, J., Beveridge, M.C.M., Byron, C.J., Clement, A., Corner, R., Costa-Pierce, B.A., Cross, S., De Wit, M., Dong, S., Ferreira, J.G., Kapetsky, J.M., Karakassis, I., Leschen, W., Little, D., Lundebye, A.K., Murray, F.J., Philip, M., Ramos, L., Sadek, S., Scott, P.C., Valle-Levinson, A., Waley, D., White, P.G., Zhu, C., (2013). Carrying capacities and site selection within the ecosystem approach to aquaculture. In: Ross, L.G., Telfer, T.C., Falconer, L., Soto, D., Aguilar-Manjarrez, J. (eds) Site selection and Carrying Capacities for Inland and Coastal Aquaculture, pp. 19-46. FAO Fisheries and Aquaculture Proceedings No. 21, Rome.
- Rothuis, A.M., Turenhout, A., Van Dujin, A., Roem, E.,
 Rurangwa, E., Katunzi, A., Shoko
 Kabagamba, J.B., (2014). Aquaculture in
 East Africa; A regional approach.
 Wageningen LE, Wageningen UR
 (University and Research centre). LE
 Report IMARESS.
- Spigel, R.H., Coulter, G.W., (1996). Comparison of hydrology and physical limnology of East African Great Lakes: Tanganyika, Malawi, Victoria, Kivu and Turkana (with reference

- to some North American Great Lakes) in : The limnology climatology and paleoclimatology of East African Lakes (ed.by T.C Johnson and E. Odada), 103-139 Gordon and Breach. Amsterdam.
- Vollenweider, R.A., Giovanardi, F., Montanari, G., Rinaldi, A., (1998). Characterization of trophic conditions of marine coastal waters with special reference to the NW Adriatic Sea: proposal for a trophic scale, turbidity and generalized water quality index. DOI:org/10.1002/(SICI) 1099.095X(199805/06) 9:3<3291 AID-ENV308>3.0<0:2-9
- WMO, (1974). Hydro-meteorological Survey of the Catchments of Lakes Victoria, Kyoga and Albert (4 vols). WMO, Geneva ISSN: 0262-6667.
- WMO, (1981). Hydro-meteorological Survey of the Catchments of Lakes Victoria, Kyoga and Mobutu Sese Seko (1 vol). WMO, Geneva ISSN: 0262-6667.
- Worm, B., Hilborn, R.J.K., Baum, T.A., Branch, (2009). Rebuilding Global Fisheries, Science 325, 578. DOI: 10.11261 science 1173146.
- Xungang, Y., Sharon, E.N., (1998). The water balance of Lake Victoria, Hydrological Sciences Journal 43(5), 789-811.